# ERRATA SHEET TENTATIVE ORDER NO. R9-2009-0001 NPDES NO. CA0107409

# WASTE DISCHARGE REQUIREMENTS AND NATIONAL POLLUTANT DISCHARGE ELIMINATION SYSTEM PERMIT FOR THE CITY OF SAN DIEGO E.W. BLOM POINT LOMA METROPOLITAN WASTEWATER TREATMENT PLANT DISCHARGE TO THE PACIFIC OCEAN THROUGH THE POINT LOMA OCEAN OUTFALL, SAN DIEGO COUNTY

The following changes have been made to Tentative Order No. R9-2009-0001. Changes below are shown in **bold and underline**/strikeout format to indicate added and removed language, respectively.

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
1	12	III.B	Based on Comment 14 (a):  Discharge through the PLOO from the Facility in excess of anmonthly average daily-flow rate of 240 MGD is prohibited.
2	47	VI.C.6.b	Based on Comment 14 (d):  The Discharger shall prepare a feasibility study that assesses behavior of the PLOO wastewater plume and means of tracking the plume. The feasibility study shall present a recommended plan for plume tracking which includes identifying recommended modifications in receiving water sampling parameters, locations, and/or sampling protocols. The feasibility study shall be submitted to the Executive Director within two years of the effective date of this Order.

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)						
3	E-21 and E-22	Table E-5 (MRP)	Based on Comment  Table E-5. Genera	,	uality Mon	itoring Req			
			Parameter	Units	Sample Type	Mi	inimum Samp Frequency		Required Analytical Test Method
					Туре	Offshore Stations	Kelp Stations	Shoreline Stations	
			Temperature	°C	Profile	1/Quarter	5/Month	Stations	1
			Salinity	ppt	Profile	1/Quarter	5/Month		1
			Dissolved Oxygen	mg/L	Profile	1/Quarter	5/Month		1
			Light Transmittance	%	Profile	1/Quarter	5/Month		1
			Chlorophyll a	m	Profile	1/Quarter	5/Month		1
			рН	units	Profile	1/Quarter	5/Month		1
			Ammonium (NH4+)	mg/L	Profile	1/Quarter	5/Month		1 <u>, 3</u>
			Visual Observations <sup>2</sup>		Visual	1/Quarter	5/Month	5/Month	
			As specified in 40 CFF Visual observations sh wind (direction and specified in 40 CFF Visual observations sh wind (direction and specified in 40 CFF visual observations sh shall be recorded. Ob sewage origin in the w a sample is collected. be noted and reported  Shall be monitored as	nall note the peed), weathe servations of rater or on the Further, the	r (e.g., cloudy water color, e beach shall nature and e	y, sunny, or rai discoloration, o be recorded. xtent of primar	ny), and tidal on the second of the second o	conditions (e.g., I turbidity, odor, r ations shall be re eation use in fede	nigh or low tide) materials of corded whenever eral waters must
4	F-47	VI.D.X.1.a (Fact Sheet)	A grid of 36 offshore times per month for the dissolved oxygen, and throughout the entire monitored at the sa	stations is he following nmonium (f water colu	g paramete <del>NH4+)</del> , and Imn <u>, with t</u> l	ers: salinity, to I chlorophyll he exceptio	emperature a. These pa n of ammo	, density, pH, arameters are nium, which	transmissivity, measured

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)	
5	E-30 and	Table E-9 (MRP)	Based on Comment 14 (e): Table E-9 replaced with the table below.	
	E-31		Reports Report Period Report Due	
			MONTHLY REPORTS Influent and effluent Solids removal/disposal Receiving water quality Tijuana cross-border emergency connection (when flowing)  Monthly By the 1 <sup>st</sup> day of 2 <sup>nd</sup> following month (e.g., March 1 for January)	
			QUARTERLY REPORTS Sludge analysis  January-March April-June July-September October-December  March 1	
			SEMI-ANNUAL REPORTS Pretreatment report January-June July-December March 1	
			ANNUAL REPORTS Pretreatment report Sludge analysis QA report Flow measurement Outfall inspection Receiving waters monitoring Kelp report  January-December April 1 April 1 April 1 July 1 July 1 July 1 July 1 October 1	

ERRATA#	PAGE	SECTION/TABLE	R	EVISION (See Responses to C	Comments for Refere	ence)	
6	51	VII.I.2.e.ii	Based on Comment 14 (i):  For all-bacterial analyses, sample dilutions should be performed so the range of values extends as follows:				
			• 2 to 12,000/100m	nl colony-forming units (C nl CFU for fecal coliforms nl CFU for enterococci		<u>iforms</u>	
			shall be reported with the and fecal) shall be those been approved in advan- enterococcus shall be th for Escherichia coli and it	elony-forming units). The ele results of the analysis. De presented in Table 1A of ele by USEPA, pursuant to lose presented in USEPA per Enterococci in Water by Med method determined by the	Detection methods 40 CFR 136, unlea 40 CFR 136. Detection Debublication EPA 60 Dembrane Filter Pro	used for colifo ss alternate me tection method: 00/4-85/076, Te ocedure, listed	rm (total ethods have is used for est Methods under 40
7	B-1	Attachment B	Based on Comment 14 ( Errata Sheet.	(I): Attachment B replaced	with the bathymet	ric map attache	ed to the
8	E-4 to E-6	Table E-1	Based on Comment 14 (m): The F station locations have been updated in Table E-1 with the information below.				
			Discharge Point Name	Monitoring Location Name	Latitude	Longitude	Depth (m)
				OFFSHORE MONITOR		1	
				F-001	32.637683	-117.240316	18 <sup>1</sup>
				F-002	32.756966	-117.272733	18 <sup>1</sup>

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)				
				F-003	32.781833	-117.272416	18 <sup>1</sup>
				F-004	32.594533	-117.26875	60 <sup>2</sup>
				F-005	32.611683	-117.26965	60 <sup>2</sup>
				F-006	32.630833	-117.2736	60 <sup>2</sup>
				F-007	32.651134	-117.279994	60 <sup>2</sup>
				F-008	32.67215	-117.283	60 <sup>2</sup>
				F-009	32.68555	-117.286316	60 <sup>2</sup>
				F-010	32.705419	-117.290658	60 <sup>2</sup>
				F-011	32.725544	-117.294632	60 <sup>2</sup>
				F-012	32.746583	-117.302066	60 <sup>2</sup>
				F-013	32.765383	-117.3072	60 <sup>2</sup>
				F-014	32.781559	-117.311423	60 <sup>2</sup>
				F-015	32.5941	-117.28645	80 <sup>3</sup>
			-	F-016	32.611833	-117.290066	80 <sup>3</sup>
				F-017	32.630016	-117.294166	80 <sup>3</sup>
			-	F-018	32.649766	-117.298333	80 <sup>3</sup>
			-	F-019	32.66785	-117.306833	80 <sup>3</sup>
				F-020	32.685416	-117.310966	80 <sup>3</sup>
				F-021	32.7038	-117.318687	80 <sup>3</sup>
				F-022	32.72273	-117.320902	80 <sup>3</sup>
				F-023	32.741883	-117.330416	80 <sup>3</sup>
				F-024	32.761216	-117.33645	80 <sup>3</sup>
				F-025	32.77895	-117.343583	80 <sup>3</sup>
				F-026	32.593766	-117.3122	98 <sup>4</sup>
				F-027	32.611783	-117.321383	98 <sup>4</sup>
				F-028	32.629287	-117.323721	98 <sup>4</sup>
			-	F-029	32.647815	-117.32493	98 <sup>4</sup>
			-	F-030	32.66567	-117.32483	98 <sup>4</sup>
				F-031	32.684668	-117.328353	98 <sup>4</sup>
			-	F-032	32.701416	-117.334166	98 <sup>4</sup>
				F-033	32.720466	-117.339916	98 <sup>4</sup>

ERRATA#	PAGE	SECTION/TABLE	REVI	SION (See Re	sponses to Commer	nts for Refere	ence)	
				F-034	3	2.7389	-117.349366	98 <sup>4</sup>
				F-035	3	2.7577	-117.363383	98 <sup>4</sup>
				F-036	32	.776783	-117.374566	98 <sup>4</sup>
9	E-7	III.A (MRP)	Based on Comment 14 (n): been updated as follows.  Monitoring Location INF-Influent monitoring is required industrial source control proceeding evaluate compliance with eaccurately characterize raw unaffected by in-plant returnmentoring shall be conduct the table below.	001 and EM red to detern ograms, to a ffluent limita wastewate n or recycle	IG-001  nine the effectivents seess the performetions. As such, in the entire seed flows or the additi	ness of pretr nance of trea ifluent moni ervice area ion of treatn	reatment and atment faciliti toring results of the treatm nent chemica	non- es, and to must ent facilities, ls. Influent
				may Cannaati	ion Monitoring of INI	E 001 and EN	IC 004	
			Table E-2. Influent and Emerge Parameter	Units	Sample Type	Minimum	Sampling uency	Required Analytical Test Method
			Flow rate	MGD	recorder/totalizer	Conti	nuous	1
			Biochemical Oxygen Demand (5-day @20°C) (BOD <sub>5</sub> )	mg/L	24-hr composite	1/Week at	<u>INF-001</u> t EMG-001	1
			Volatile Suspended Solids	mg/L	24-hr composite	1/Week at	<u>INF-001</u> t EMG-001	1
			Total Dissolved Solids (TSS)	mg/L	24-hr composite		<u> INF-001</u> t EMG-001	1
			Temperature	°C	grab		INF-001 EMG-001	1
			Floating Particulates	mg/L	24-hr composite		INF-001 EMG-001	1
				TA	BLE A PARAMETER	S		
			Oil and Grease	mg/L	grab		INF-001 EMG-001	1

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)				
			Total Suspended Solids	mg/L	24-hr composite	1/Week at EMG-001	1
			Settleable Solids	ml/L	grab	1/Day <u>at INF-001</u> 1/Week at EMG-001	1
			Turbidity	NTU	grab	1/Day <u>at INF-001</u> 1/Week at EMG-001	1
			pН	units	grab	1/Day <u>at INF-001</u> <u>1/Week at EMG-001</u>	1
10	16	Table 10	Based on Comment 14 (o):	The portion	of Table 10 sho	own below has been up	dated as follows.
			Chromium, Total Recoverable	μg/L			3.9E+07
			(III) <sup>2</sup>	lbs/day			6.7E+07
11	E-8 E-11	Table E-2 (MRP) Table E-3	Based on Comment 14 (o): follows.  Chromium (III), Total Recoverable <sup>2</sup> Based on Comment 14 (o):	The portion	of Table E-2 sh		pdated as
		(MRP)	Chromium (III) <sup>2</sup>	μg/L	24-hr composite	2 1/Month	1
13	F-36	Table F-16	Based on Comment 14 (o):				
13	F-30	(Fact Sheet)	based on Comment 14 (0).				

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
			lbs/day 6.7E+07
14	E-14	V.A.1 (MRP)	Based on Comment 14 (p):  The Discharger shall conduct monthly chronic toxicity tests on 24-hour composite effluent samples. Once each calendar year, at a different time of year from the previous years, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct three toxicity tests using a fish, an invertebrate, and an alga species; the Discharger shall then continue to conduct routine monthly toxicity testing using the single, most sensitive species. For the initial three suites of chronic toxicity tests, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct toxicity tests using a fish, an invertebrate, and an alga species. After the initial screening period, the Discharger shall conduct routine monthly toxicity testing using the most sensitive species. Every other year, the Discharger shall re-screen at a different time from the prior years. Re-screening can be limited to one month, if results are the same as the previous three-month screening. However, if results of the re-screening are different, then the Discharger shall conduct two additional months of re-screening to determine the most sensitive species and then conduct routine monthly toxicity testing using the most sensitive species.
15	E-17	V.A.3.j (MRP)	Based on Comment 14 (r):  pH drift during the toxicity test may contribute to artifactual toxicity when pH-dependent toxicants (e.g., ammonia, metals) are present in an effluent. To determine whether or not pH drift during the toxicity test is contributing to artifactual toxicity, the Discharger shall conduct three sets of parallel toxicity tests, in which the pH of one treatment is controlled at the pH of the effluent and the pH of the other treatment is not controlled, as described in Section 11.3.6.1 of the test methods manual, Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms (EPA/821/R-02/013, 2002). Toxicity is confirmed to be artifactual and due to pH drift when no toxicity above the chronic toxicity effluent limit is observed in the treatments controlled at the pH of the effluent. If toxicity is

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
			confirmed to be artifactual and due to pH drift, then, following written approval by the Executive Officer and USEPA, the Discharger may use the procedures outlined in Section 11.3.6.2 of the test methods manual to control sample pH during the toxicity test.
16	E-20	V.B.3 (MRP)	Based on Comment 14 (r): The text below has been deleted from Section V.A.3.j in the MRP and added as Section V.B.3.k in the MRP.  k. pH drift during the toxicity test may contribute to artifactual toxicity when pH-dependent toxicants (e.g., ammonia, metals) are present in an effluent. To determine whether or not pH drift during the toxicity test is contributing to artifactual toxicity, the Discharger shall conduct three sets of parallel toxicity tests, in which the pH of one treatment is controlled at the pH of the effluent and the pH of the other treatment is not controlled, as described in Section 11.3.6.1 of the test methods manual, Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms (EPA/821/R-02/013, 2002). Toxicity is confirmed to be artifactual and due to pH drift when no toxicity above the toxicity effluent limit is observed in the treatments controlled at the pH of the effluent. If toxicity is confirmed to be artifactual and due to pH drift, then, following written approval by the Executive Officer and USEPA, the Discharger may use the procedures outlined in Section 11.3.6.2 of the test methods manual to control sample pH during the toxicity test.
17	E-17	V.A.4.a (MRP)	Based on Comment 14 (s):  A full laboratory report for all toxicity testing shall be submitted as an attachment to the DMR for the month in which the toxicity test was conducted and shall also include: the toxicity test results <a href="mailto:asin-NOEC">asin-NOEC</a> ; TUc = 100/NOEC;

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
18	E-18	V.B.1 (MRP)	Based on Comment 14 (t):  The Discharger shall conduct semi-annual acute toxicity tests on 24-hour composite effluent samples. Once each calendar year, at a different time of year from the previous years, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct two toxicity tests using a fish and an invertebrate species; the Discharger shall then continue to conduct routine semi-annual toxicity testing using the single, most sensitive species. For the initial three suites of acute toxicity tests, the Discharger shall split a 24-hour composite effluent sample and concurrently conduct toxicity tests using a fish and an invertebrate. After the initial screening period, the Discharger shall conduct routine semi-annual toxicity testing using the most sensitive species. Every other year, the Discharger shall re-screen at a different time from the prior years. Re-screening can be limited to one month, if results are the same as the previous three-month screening. However, if results of the re-screening are different, then the Discharger shall conduct two additional months of re-screening to determine the most sensitive species and then conduct routine semi-annual toxicity testing using the most sensitive species.
19	E-25	VIII.A.3 (MRP)	Based on Comment 14 (u):  Twice per year (January and July), sediment samples for benthic infauna community structure shall be collected from the offshore sediment monitoring locations specified in Table E-1, which consists of 12 primary stations and an additional 10 secondary stations. Two replicate samples shall be taken using a 0.1 square meter modified Van Veen grab sampler. These samples shall be separate from those collected for grain size and chemistry. The samples shall be sieved using a 1.0-mm mesh screen. The benthic organisms retained on the sieve shall be fixed in 105 percent buffered formalin and transferred to at least 70 percent ethanol within two to seven days for storage. All retained benthic infauna organisms shall be counted and identified to as low a taxon as possible. This enumeration and identification of organisms continues to use the historical database developed by the Discharger.

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
			Analysis of benthic community structure shall include determination of the number of species, number of individuals per species, and total numerical abundance present. The following parameters shall be <a href="calculated for each grab sample and summarized by station as appropriate-summarized for each station">calculated for each grab sample and summarized by station as appropriate-summarized for each station:</a> a. Average number of species (species richness) per 0.1 m²; b. Total numerical abundance; d. Infaunal trophic index (ITI); e. Benthic response index (BRI); f. Swartz' 75% dominance index; g. Shannon-Weiner's diversity index (H'); and h. Pielou evenness (J') a. Number of species per 0.1m² (species richness); b. Total (cumulative) number of species per station; c. Total numerical abundance; d. Benthic response index (BRI); e. Swartz's 75% dominance index; f. Shannon's diversity index (H'); and g. Pileou's evenness index (J').
20	E-26	VIII.A.4 (MRP)	Based on Comment 14 (v):  Chemical analyses of fish tissues shall be performed annually on target species collected at or near the trawl and rig fishing stations. The various stations are classified into zones for the purpose of collecting sufficient numbers of fish for tissue analyses. Trawl Zone 1 represents the nearfield zone, defined as the area within a 1-km radius of stations SD-010 and/or SD-012; Trawl Zone 2 is considered the northern farfield zone, defined as the area within a 1-km radius of stations SD-013 and/or SD-014; Trawl Zone 3 represents the LA-5 disposal site zone, and is defined as the area centered within a 1-km radius of station SD-008; Trawl Zone 4 is

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
			considered the southern farfield zone, and is defined as the area centered within a 1-km radius of station SD-007. The two rig fishing stations also represent two distinct zones. Rig feishing zeone 1 is the nearfield area centered within a 1-km radius of Station RF-001; feight feishing zeone 2 is considered the farfield area centered within a 1-km radius of station RF-002. There are no depth requirements for these six zones with regards to the collection of fishes for tissue analysis.
			Liver tissues shall be analyzed semiannually (January and July)annually (i.e., during October) from fishes collected in each of the above four trawl zones.—Each trawl station may be trawled up to a maximum of five times in order to acquire sufficient numbers of fish for composite samples within a zone; trawls subsequent to the initial community trawl discussed above (i.e., trawls 2-5/site) may occur anywhere within a defined zone. No more than a maximum of five 10-minute (bottom time) trawls shall be required per zone in order to acquire sufficient numbers of fish for composite samples; these trawls may occur anywhere within a defined zone. Three replicate composite samples shall be prepared from each trawl zone, with each composite consisting of tissues from as least three individual fish of the same species collected within a zone. These liver tissues shall be analyzed for the presence and concentrations of lipids, PCB (congeners), chlorinated pesticides, and the following three metals: mercury, arsenic and selenium. The species of fish targeted for tissue analysis at from the trawl sites shall be primarily flatfish, and include including, but not limited to, the longfin sanddab (Citharichthys xanthostigma) and the Pacific sanddab (Citharichthys sordidus). If sufficient numbers of these primary target-species are not present in a zone, secondary candidate species such as other flatfish or rockfish may be collected as necessary.
			Rig fishing shall be performed Muscle tissues shall be analyzed annually (i.e., during October) to monitor the uptake of pollutants in fish species which are from fishes collected in each of the above two rig fishing zones in order to monitor the uptake of pollutants in species and tissues that are consumed by humans. These fishspecies shall be representative of those caught by recreational and or commercial fishery activities in the region. All fish shall be collected by hook and line or by setting baited lines or traps within the two rig fishing zones described above. The species targeted for analysis at the rig fishing sites shall be primarily rockfish, which may include, but are not limited to and include the vermilion

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
			rockfish ( <i>Sebastes miniatus</i> ) and the copper rockfish ( <i>Sebastes caurinum</i> ). If sufficient numbers of these primary fish-species are not present or cannot be caught in a particular zone, othersecondary target species (e.g., rockfish, scorpionfish) may be collected and analyzed as necessary. Three replicate composite samples of the target species shall be obtained from each zone, with each composite consisting of a minimum of three individual fish. Muscle tissues shall be removed from the composites and chemically analyzed for the presence and concentrations of lipids, PCB (congeners), chlorinated pesticides, and the following nine metals: arsenic, cadmium, chromium, copper, lead, mercury, selenium, tin and zinc.
21	F-48	VI.D.1.d (Fact Sheet)	Based on Comment 14 (v):  Twice each year, in January and July, eEpibenthic trawls at four trawl zone stations are used to assess the structure of demersal fish and megabenthic invertebrate communities and to evaluate compliance with narrative water quality standards in the Ocean Plan. Semiannually, in January and July, eChemical analyses of fish tissues are performed on target species colleted at the four trawl zone stations and two rig fishing stations. Species targeted for analysis are selected based on their ecological and/or commercial importance. Liver tissue is monitored at trawl stations to assess general fish health. Muscle tissue is monitored at rig fishing stations annualy, in October, to assess the uptake of pollutants in fish species commonly consumed by humans in the region. Fish tissues are monitored for lipids, metals, PCBs, and chlorinated pesticides.
22	E-30	IX.B.1 (MRP)	Based on Comment 14 (x):  At any time during the term of this permit, the State or Regional Water Board may notify the Discharger to electronically submit Self-Monitoring Reports (SMRs) using the State Water

ERRATA#	PAGE	SECTION/TABLE	REVISION (See Responses to Comments for Reference)
			Board's California Integrated Water Quality System (CIWQS) Program Web site (http://www.waterboards.ca.gov/ciwqs/index.html). Until such notification is given, the Discharger shall submit hard copy SMRs. For this purpose, a hard copy signed penalty of perjury statement accompanying a CD with a single file in PDF format (including the certification specified in Section V.B. 5 of Attachment D) shall qualify as a hard copy SMR. The CIWQS Web site will provide additional directions for SMR submittal in the event there will be service interruption for electronic submittal.
23	-	-	Based on Comment 14 (aa): Typographical errors and other minor corrections to the wording in the tentative order have been or will be made prior to sending out the final version.
24	43	VI.C.5.c.v	Based on a recommendation by the United States Environmental Protection Agency (USEPA) Region 9 Clean Water Act Compliance Office:  By AprilMarch 1 of each year, the Discharger shall submit an annual report to the Regional Water Board; USEPA Region 9; the State Water Board, Division of Water Quality, Regulations Unit; and the San Diego County Department of Health Services, Hazardous Materials Division, describing its pretreatment activities over the previous calendar year.
25	C-1	Attachment C	Attachment C figures replaced with schematic diagrams (including disinfection) attached to the Errata Sheet.

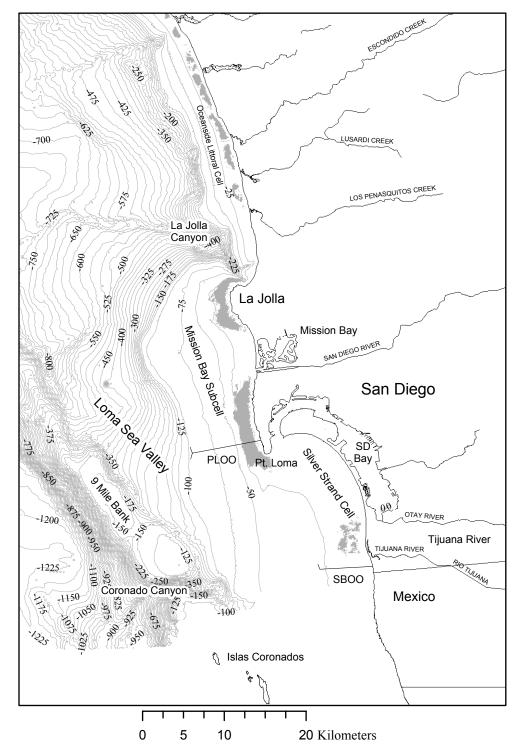
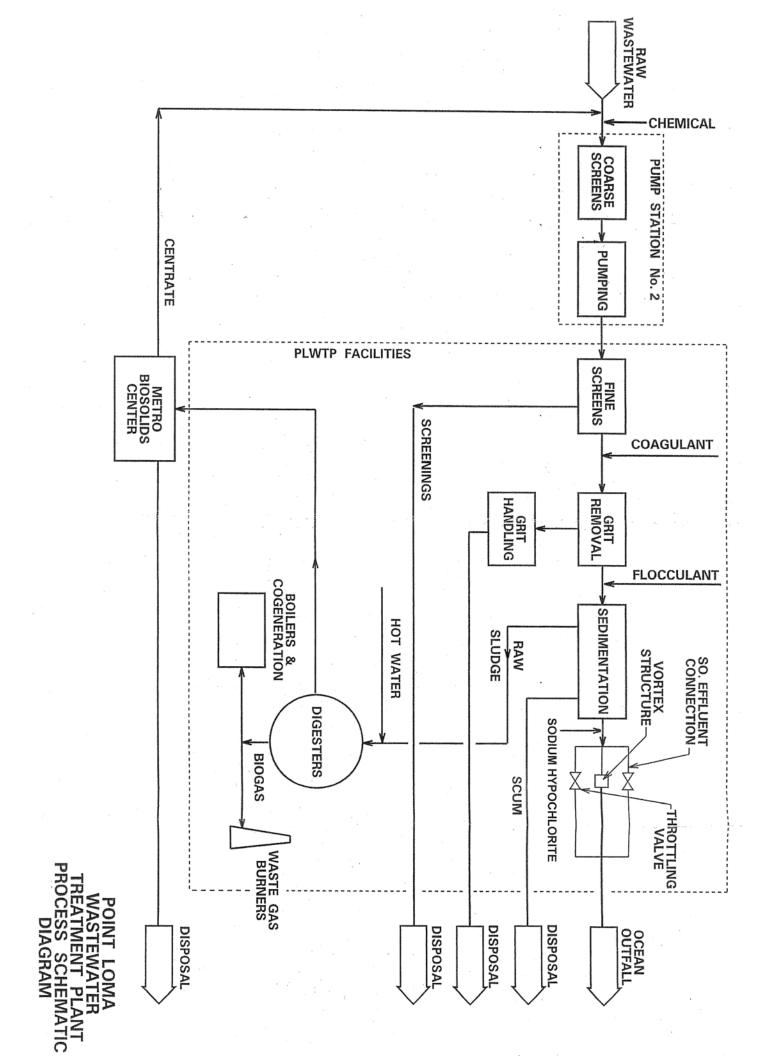


Figure 1. Map of marine shelf of San Diego County. Bathymetric units are meters. Locations of littoral cells, submarine canyons, outfalls (PLOO and SBOO), rivers, and Kelp Forests (shaded areas close to shore) are indicated.



METRO SYSTEM PROCESS SCHEMATIC